

Short Report

Most Azole-Susceptible Isolates of *Aspergillus fumigatus* in Hokkaido, Japan are Clustered with Worldwide Strains That Do Not Have Tandem Repeats in *Cyp51A* Promoter

Takuya Fujiwara¹, Megumi Itoh¹, Naoya Matsumoto^{2,3}, Yusuke Koshizaki⁴, Syunpei Saitou⁴, Kazutaka Yamada², and Takahito Toyotome^{1,5,6}

¹ Department of Veterinary Medicine, Obihiro University of Agriculture and Veterinary Medicine

² School of Veterinary Medicine, Azabu University

³ Noboribetsu Marine Park Nixe

⁴ JA-Hokkaidoukouseiren Obihiro Hospital

⁵ Diagnostic Center for Animal Health and Food Safety, Obihiro University of Agriculture and Veterinary Medicine

⁶ Medical Mycology Research Center, Chiba University

ABSTRACT

In this study, we analyzed *Aspergillus fumigatus* short tandem repeat patterns of 106 strains isolated from the outdoor air, clinical specimens, and king penguins (*Aptenodytes patagonicus*) with aspergillosis in Japan, and compared them with those of 668 strains from AfumID (including six isolates from Japan). The results showed that the isolates were classified into three major groups. Group II contained most of the azole-resistant strains with 34- and 46-bp tandem repeats in *cyp51A* promoter. As in our previous study, OKH50 and Env1 strains were classified in Group II. Most of the azole-susceptible strains obtained in Japan were classified in Group III.

Key words : *Aspergillus fumigatus*, azole-resistant, short tandem repeat

Our previous studies demonstrated that *Aspergillus fumigatus* strains OKH50 and Env1 with 34-bp tandem repeats (TR₃₄) in the *cyp51A* promoter isolated respectively from a patient and an environmental sample in Japan had identical short tandem repeat (STR) patterns^{1,2}. The analysis of the STR pattern of 34 strains in the previous study¹ showed that OKH50, Env1, and azole-resistant strains with TR₃₄ in the *cyp51A* promoter isolated in Europe, Middle East, India, and Australia were closely related^{1,2}. However, only OKH50 and nine strains isolated in Japan were examined in our previous study¹. Therefore, the relation between OKH50 and Env1 with other *A. fumigatus* strains is not fully understood. In this study, we analyzed *A. fumigatus* STR patterns of 89 strains isolated from clinical specimens and one strain, Env1, isolated from the environment in Obihiro, Japan. The clinical isolates included several sets of strains obtained from the same

patients. Fourteen strains isolated from the captive environment of king penguins (*Aptenodytes patagonicus*) in Noboribetsu of Japan and two strains isolated from king penguins with aspergillosis in Noboribetsu of Japan were also included in the analysis. We then compared them with STR patterns of 668 strains isolated around the world.

The strains used in this study are shown in Supplementary Table 1. Fungal species of these strains were identified with the sequences of internal transcribed spacer regions (ITS1 and 2) and partial nucleotide sequences of β -tubulin, *rodA*, and calmodulin genes as described previously². For this study, we examined 89 clinical isolates, which had been isolated at a hospital in Obihiro City, including 67 isolates from our earlier research³. At Noboribetsu City, Hokkaido in Japan, 3 strains from the air and 11 strains isolated from swabs swiped at the surface of the aquarium facility for captive king penguins were

Address for correspondence: Takahito Toyotome, PhD

Department of Veterinary Medicine, Obihiro University of Agriculture and Veterinary Medicine, Nishi 2-11, Inada-cho, Obihiro, Hokkaido 080-8555, Japan

Received: 5 June 2023, Accepted: 3 August 2023

E-mail: tome@obihiro.ac.jp

collected⁴). Two strains from king penguins with aspergillosis used for this study were isolated as described in our earlier studies^{5,6}. Genomic DNA from *A. fumigatus* strains was prepared as described elsewhere⁷. To amplify STR loci, we used primers described by de Valk⁸ for this study using the usual techniques. Fragment analysis and sizing analysis were performed with a 3730xl DNA Analyzer (Thermo Fisher Scientific Inc., Waltham, MA) by Fasmac Co., Ltd. (Kanagawa, Japan). For genetic analysis using the STR pattern, the poppr ver. 2.9.3^{9,10} package of R software¹¹ was used to calculate the average Bruvo's distance over all loci in a population; UPGMA was used for the clustering. In addition to the 104 strains analyzed for STR in this study, we included STR patterns of OKH50 and Env1 as previously reported^{1,2} and 668 strains from AfumID (<https://afumid.shinyapps.io/afumid/>)¹² in the analysis (Supplementary Table 2).

The 106 strains obtained in Japan and 668 strains obtained from AfumID were classified into three major groups on the basis of the STR patterns (Supplementary Fig. 1). Group III contained only 11 of 180 TR₃₄-type azole-resistant strains (Table 1). From Japan, 28 and 84 isolates were included in Groups II and III, respectively (Supplementary Fig. 1). The *A. fumigatus* strains OKH50 and Env1, which were TR₃₄-type azole-resistant isolates reported previously^{1,2}, were included in Group II, which is consistent with our earlier result. The other 26 Japanese isolates were azole-susceptible strains. As shown by Hagiwara¹³ and Nakano¹⁴, TR₃₄- and TR₄₆-type azole-resistant strains were isolated from imported plant bulbs such as *Tulipa* spp. and *Gladiolus* spp. Various ornamental flowering plants have been imported from Europe to Japan for over 100 years¹⁵. The Japanese isolates included in Group II might have originated from the Eurasian Continent before the global spread of TR₃₄- and TR₄₆-type strains and might have colonized the domestic environment. Similar to the global expansion of pathogenic chytrid fungi¹⁶, international trade of agricultural crops planted in the soil might be the cause of the disturbance of native species or strains and the environment.

Minimum inhibitory concentrations (MICs) of four antifungals against isolates from the same patients with identical STR patterns were determined with a broth microdilution method described earlier². The data are shown in Supplementary Table 3. OKH186 showed quite slow growth. The strain was isolated 28 months after OKH131 was isolated. The MIC values of voriconazole and itraconazole determined at 48 hrs after inoculation were reduced to 1/8 or less than MICs against OKH131 (Supplementary Table 3), suggesting that the strain had changed qualitatively to survive in the host during chronic infection. OKH185 showed four- and eight-times higher MICs for itraconazole and voriconazole, respectively, compared to OKH141, which was isolated 18 months before OKH185 (Supplementary Table 3). The sequence of *cyp51A* gene and the promoter showed no mutation or tandem repeat. However,

Table 1. Numbers of *Aspergillus fumigatus* strains in each group shown in Supplementary Fig. 1.

Group	Non-TR strains	TR ₃₄ -type	TR ₄₆ -type
I	2 (0)	2 (0)	0 (0)
II	260 (26)	168 (2)	83 (0)
III	248 (84)	11 (0)	0 (0)
Total	510 (110)	181 (2)	83 (0)

The numbers of Japanese isolates are shown in parentheses.

TR: tandem repeats

the comparison of *hmg1* gene between OKH141 strain and OKH185 strain showed a point mutation resulting in an amino acid substitution at the 305th position from serine to proline. The mutation reported in earlier publications^{17,18} contributed to the decrease of azole susceptibility¹⁸.

Of the 16 isolates from the environment and king penguins with aspergillosis in an aquarium, 9 were clustered in a branch (Supplementary Fig. 1), suggesting that they are domestic STR types and that the king penguins had acquired the causative strains from the environment. However, the cluster also contains many strains isolated from Obihiro City. Further research to clarify geographical differences is warranted.

Acknowledgments

This work was supported by JSPS Grants-in-Aid for Scientific Research (C) Numbers 17K08095 and 21K05931, and internal funding of Obihiro University of Agriculture and Veterinary Medicine (Hokkaido, Japan). We thank Prof. Matthew C. Fisher and colleagues for allowing us to use STR data from AfumID.

Conflicts of interest

None.

References

- 1) Toyotome T, Hagiwara D, Kida H, Ogi T, Watanabe A, Wada T, Komatsu R, Kamei K: First clinical isolation report of azole-resistant *Aspergillus fumigatus* with TR₃₄/L98H-type mutation in Japan. *J Infect Chemother* **23**: 579-581, 2017.
- 2) Onishi K, Muhammad Sarumoh B, Hagiwara D, Watanabe A, Kamei K, Toyotome T: Azole-resistant *Aspergillus fumigatus* containing a 34-bp tandem repeat in *cyp51A* promoter is isolated from the environment in Japan. *Med Mycol J* **58**: E67-E70, 2017.
- 3) Toyotome T, Saito S, Koshizaki Y, Komatsu R, Matsuzawa T, Yaguchi T: Prospective survey of *Aspergillus* species isolated from clinical specimens and their antifungal susceptibility: a five-year single-center study in Japan. *J Infect Chemother* **26**:

- 321-323, 2020.
- 4) Matsumoto N, Itoh M, Yamada K, Toyotome T: Detection of *Aspergillus fumigatus* in the captive environment and preventive measures leading to the eradication of penguin aspergillosis in an aquarium. *Jpn J Zoo Wildl Med* **25**: 101-107, 2020. [Article in Japanese]
 - 5) Yamada K, Toyotome T, Matsumoto N, Itoh M: Autopsy imaging for aspergillosis in King Penguin, an economically valuable zoo animal. *J Vet Med Sci* **82**: 373-375, 2020.
 - 6) Itoh M, Toyotome T, Matsumoto N, Okamoto M, Watanabe KI, Yamada K: Characteristic imaging findings of the respiratory system in penguins with suspected aspergillosis in an aquarium. *J Vet Med Sci* **82**: 1260-1266, 2020.
 - 7) Bok JW, Keller NP: Fast and easy method for construction of plasmid vectors using modified quick-change mutagenesis. *Methods Mol Biol* **944**: 163-174, 2012.
 - 8) de Valk HA, Meis JFGM, Curfs IM, Muehlethaler K, Mouton JW, Klaassen CHW: Use of a novel panel of nine short tandem repeats for exact and high-resolution fingerprinting of *Aspergillus fumigatus* isolates. *J Clin Microbiol* **43**: 4112-4120, 2005.
 - 9) Kamvar ZN, Tabima JF, Grünwald NJ: Poppr: an R package for genetic analysis of populations with clonal, partially clonal, and/or sexual reproduction. *PeerJ* **2**: e281, 2014.
 - 10) Kamvar ZN, Brooks JC, Grünwald NJ: Novel R tools for analysis of genome-wide population genetic data with emphasis on clonality. *Front Genet* **6**: 208, 2015.
 - 11) R Core Team R: A language and environment for statistical computing. 2022. <https://www.R-project.org/>
 - 12) Sewell TR, Zhu J, Rhodes J, Hagen F, Meis JF, Fisher MC, Jombart T: Nonrandom distribution of azole resistance across the global population of *Aspergillus fumigatus*. *mBio* **10**: e00392-19, 2019.
 - 13) Hagiwara D: Isolation of azole-resistant *Aspergillus fumigatus* from imported plant bulbs in Japan and the effect of fungicide treatment. *J Pestic Sci* **45**: 147-150, 2020.
 - 14) Nakano Y, Tashiro M, Urano R, Kikuchi M, Ito N, Moriya E, Shirahige T, Mishima M, Takazono T, Miyazaki T, Izumikawa K: Characteristics of azole-resistant *Aspergillus fumigatus* attached to agricultural products imported to Japan. *J Infect Chemother* **26**: 1021-1025, 2020.
 - 15) Kurashige Y: Meiji kara Heisei wo irodoru hitome 30-oku yen no Tulip. Sakata Seed Corporation, 2016. https://sakata-tsushin.com/yomimono/tokushu/20160915_003848.html [in Japanese]
 - 16) Fisher MC, Pasmans F, Martel A: Virulence and pathogenicity of chytrid fungi causing amphibian extinctions. *Annu Rev Microbiol* **75**: 673-693, 2021.
 - 17) Takeda K, Suzuki J, Watanabe A, Arai T, Koiwa T, Shinfuku K, Narumoto O, Kawashima M, Fukami T, Tamura A, Nagai H, Matsui H, Kamei K: High detection rate of azole-resistant *Aspergillus fumigatus* after treatment with azole antifungal drugs among patients with chronic pulmonary aspergillosis in a single hospital setting with low azole resistance. *Med Mycol* **59**: 327-334, 2020.
 - 18) Rybak JM, Ge W, Wiederhold NP, Parker JE, Kelly SL, Rogers PD, Fortwendel JR: Mutations in *hmg1*, challenging the paradigm of clinical triazole resistance in *Aspergillus fumigatus*. *mBio* **10**: e00437-19, 2019.